CAF2020 Paper No. P-5-2-42

Navarro H, Navarro S (2021) Efficacy of controlling two stored cashew nut insects in 1%  $O_2$  and 99%  $N_2$  at 43°C. Pp. 153-159. In: Jayas DS, Jian F (eds) Proceedings of the 11<sup>th</sup> International Conference on Controlled Atmosphere and Fumigation in Stored Products (CAF2020), CAF Permanent Committee Secretariat, Winnipeg, Canada.

# Efficacy of controlling two stored cashew nut insects in 1% O<sub>2</sub> and 99% N<sub>2</sub> at 43°C

Hagit Navarro\* and Shlomo Navarro

Green Storage Ltd., Argaman 5, Rishon Letsion, 7570905, Israel. \*Corresponding author's email: hnavarro@green-storage.co.il

# Abstract

The efficacy of controlling all life stages of storage insects Tribolium castaneum (Herbst) and Oryzaephilus surinamensis (L.) were tested at 43°C and in 1% O2 and 99% N2 for 48 h of exposure time. For each species and each life stage, four replicates (each replicate consisted of 50 insects) were conducted. Two replicates for each species and each life stage served as control. Two control replicates were compared with treatment, one replicate served as control at 28°C and  $60\pm5\%$  RH and another served as control at 43°C and  $60\pm5\%$ RH. All tests were carried out by introducing insects to 100 g of cashew nuts in gastight 1 L containers. At the beginning and the end of each exposure time O<sub>2</sub> percent was measured. Insect counting was done immediately after exposure treatment and after 7-10 days of incubation at 28°C and 60±5% RH. Cashew nuts obtained were in equilibrium with 56.4% RH at 16.9°C. Increased mortality was observed due to exposure to 43°C in air (control) compared to exposure to 28°C. Adult stage of O. surinamensis was the most tolerant one with 99.5 % mortality after 48 h treatment. Larval and pupal stages could not survive the treatment and 100% mortality was recorded. Although in the egg stage in both species some larvae hatch was observed, they could not survive more than 24 h after the treatment. Average egg mortalities were counted as 97.5% for T. castaneum and 98% for O. surinamensis, the young hatched larvae were found dead immediately after treatment. The pupal and larval stages of both species were more susceptible to the treatment rather than the adult and egg stages. Treating both stored product insects, T. castaneum and O. surinamensis, at 43°C in 1% O<sub>2</sub> and 99% N<sub>2</sub> atmosphere for 48 h resulted in very effective control treatment. The survival rate was less than 0.5% of adult of O. surinamensis.

Keywords: Nitrogen atmosphere, Cashew nuts, Tribolium castaneum, Oryzaephilus surinamensis, High temperature

# Introduction

Cashew (*Anacardium occidentale* L.) belongs to the family Anacardiaceae and is a native of Brazil. Around the 16<sup>th</sup> century, it reached the Far East where today, Vietnam serves as the largest supplier of cashew nuts to the international market. Cashew is a high economic value crop and is earning considerable foreign exchange for the country. In 2012, about 220,000 tonnes of cashew kernels were exported, with a turnover of over US\$1.45 billion (Thai, 2012).

Since the harvest period is short, during storage, cashew nuts must be protected from insect infestation and weight loss. In most countries, food commodities fumigated with phosphine and subsequently aerated are considered to be phosphine free with no regulations attached. The US Food Quality Protection Act (FQPA), which became law in the US in 1996, has set a tolerance for phosphine of 0.01 ppm in processed food stuffs. This is well below the detection level available to most laboratories (Donahaye, 2000). As concerns about the safety of our food supply increases along with concerns about the impact of agricultural chemicals on our environment and resistant strains of stored product pests to phosphine increase, the development of nonchemical quarantine treatments to meet export requirements become increasingly necessary. Moreover, the increase in consumer demand for organic commodities in recent years has increased.

Therefore, the use of Modified or Controlled Atmospheres (MA/CA) offers a safe and environmentally benign alternative to the use of conventional residue-producing chemical fumigants for controlling insect pests that attack stored grains, oilseeds, processed commodities, and packaged foods (Navarro, 2006) and is increasing in general, and in particular for niche markets such as the treatment of organic commodities. The pre-condition for successful MA treatment is a tight hermetic seal. Sealing techniques (Andrews et al., 1994), and methods of verifying the seal (Navarro, 1999) are well developed, as are the application procedures (Navarro and Donahaye, 1990; Annis and van S. Graver 1991). Modified or Controlled Atmospheres to control insects can be obtained using nitrogen (N<sub>2</sub>), provided that a hypoxic atmosphere of  $\leq 1\%$ O<sub>2</sub> can be maintained. Generally, the lower the oxygen level, the higher the mortality. For effective control, the O<sub>2</sub> level should be <3% and preferably <1% if a rapid kill is required (Navarro, 1978).

Insect mortality increases more rapidly as temperatures rise and their metabolism speeds up. Cool temperatures slow rates of mortality while lower relative humidity hastens toxic effects, notably in high CO<sub>2</sub> atmospheres because of desiccation of insects (Banks and Fields, 1995).

Sakka et al. (2020) investigated the use of MA on life stages of several storage pests at 40°C for 2.5 d in commercial nitrogen chambers with phosphine susceptible and resistant populations of *Oryzaephilus surinamensis* (L.), *Tribolium castaneum* (Herbst) and *Sitophilus oryzae* (L.). Complete mortality of all life stages of the tested pests was obtained with negligible progeny of *O*. *surinamensis* ( $0.3\% \pm 0.3$ ) (Sakka et al., 2020). While Athanassiou et al. (2017) compared insect mortality when nitrogen (1% O<sub>2</sub>) was applied at 25°C, high insect mortality levels of *Tribolium confusum* Jacquelin du Val, *Ephestia elutella* (Hübner) and *O. surinamensis* were noted. However, in most cases there were a number of insects that survived the nitrogen treatment. In contrast, complete control was achieved at 38–43°C for all insect species and life stages tested, with the exception of *T. confusum* larvae (Athanassiou et al., 2017). Therefore, the objective of the present study was to evaluate the effect of 99% N<sub>2</sub> nitrogen treatment to achieve 1% O<sub>2</sub> on two storage pests at 43°C in a short exposure time on cashew nuts.

# Materials and methods

# Control of Temperature and $O_2$ level within atmosphere of 1% $O_2$

Insects were exposed to 1% O<sub>2</sub> in 1-L glass jars equipped with two 1/16" i.d. copper tubes soldered to the lid and seal was reassured by applying the half-time pressure decay test to hold a pressure of 6 to 3 mm H<sub>2</sub>O for at least 5 min (Navarro, 1999). The temperature and RH in the gastight 1-L volume jars holding 200 g of cashew nuts were tested using data loggers (Elitech RC-4HC, Elitech,

London, UK). The outer ends of the tubes were connected to T-type valves. An electrolytic sensor type oxygen monitor (Oxycheck Bishop, UK) equipped with an internal pump delivering 500 mL/min gas sample was used for monitoring oxygen levels in the jars. To achieve the initial target oxygen concentration, the gastight jar containing 100 g cashew nuts and the insects was purged using N<sub>2</sub> at about 1000 mL/min until the O<sub>2</sub> concentration dropped to about 0.9% in the jar. For testing the gas concentration, the two T-type valves were kept at a position so that the gas flow would be from the O<sub>2</sub> monitor to the exhaust port of the jar. In that position the T-type valves were switched simultaneously to the ports to provide a closed loop gas flow between the gastight jar and the monitor. After 24 h and at the end of the exposure, readings were taken to ensure the gas concentration was maintained during the exposure period of 48 h. Oxygen concentration was measured three times; at the beginning, after 24 h and at the end of each treatment (48 h). Equilibrium relative humidity of 500 g cashew nuts was tested at the beginning of each trial using a Novasina RH monitor (Hygro Mate, Novasina, Switzerland).

#### Insects

Populations of storage insects *T. castaneum* and *O. surinamensis* were reared at the Green Storage (GS) Ltd. laboratories at 28°C and  $60\pm5\%$  RH. The efficacy of controlling all life stages of these two species at 43°C and 99% N<sub>2</sub> for 48 h of exposure time was tested (Fig. 1). There were four replicates for each species and each life stage (eggs, larvae, pupae and adults), and each replicate consisted of 50 insects. Two replicates from each species and each life stage served as control. One replicate served as control at 28°C and  $60\pm5\%$  RH and the other replicate served as control at 43°C and  $60\pm5\%$  RH. All tests were carried out by exposing insects to 100 g of cashew nuts in gastight 1 L containers (except the control which was not gastight).

Insect counting was done immediately after each treatment and after 7-10 d of incubation at  $28^{\circ}$ C and  $60\pm5\%$ . The reason for the delayed counts was to make ensure there were no surviving insects. All hatched eggs were considered as alive.

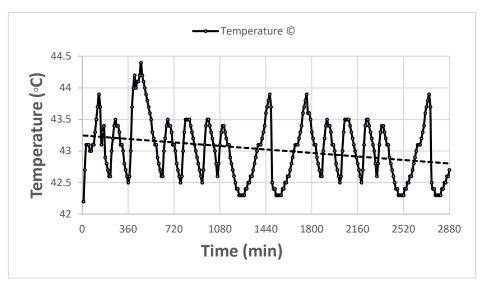


Fig. 1: Temperature obtained within the 1 L container containing 200 g of cashew nuts during a 48-h exposure time at a target temperature of 43°C.

### Results

Equilibrium relative humidity of 500 g cashew nuts was 56.4% at 16.9°C. Figures 2-3 describe the temperature and RH maintained in the 1 L container during a 48-h exposure. Average temperature was 43.0°C (Fig. 1) and the average ERH was 58.7 % (Fig. 2).

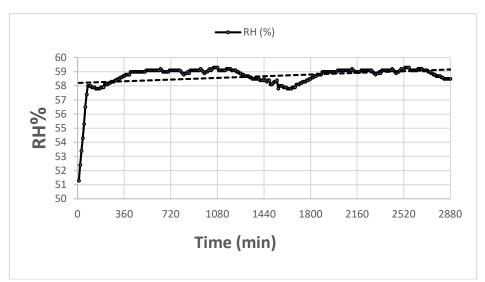


Fig. 2: Relative humidity (%) obtained in the 1 L container containing 200 g of cashew nuts during a 48-h exposure.

Table 1 shows the mortality of all life stages of *T. castaneum* and *O. surinamensis* after 48 h of exposure to 1% O<sub>2</sub> at 43°C. All larvae were dead during counting (immediately after exposure time).

Average temperature (°C)	43		43		28	
	Average O <sub>2</sub> (%)	Average mortality (%)	O2 (%)	Average mortality (%)	Average O <sub>2</sub> (%)	Average mortality (%)
Tribolium castaneum						
Adults	1.30	100.0	20.9	2.0	20.9	0.0
Pupae	1.05	100.0	20.9	42.0	20.9	0.0
Larvae	1.20	100.0	20.9	94.0	20.9	22.0
Eggs	2.35	100.0	20.9	98.0	20.9	76.0
Oryzaephilus surinamensi	5					
Adults	1.75	99.6	20.9	76.9	20.9	6.0
Pupae	1.25	100.0	20.9	90.9	20.9	62.5
Larvae	1.33	100.0	20.9	72.0	20.9	59.1
Eggs	1.45	100.0	20.9	93.0	20.9	48.0

Table 1. Oxygen concentration (%) and mortality (%) of egg, larva, pupa and adult of
Tribolium castaneum and Oryzaephylus surinamensis after 48 h of exposure to 1%
O <sub>2</sub> at 43°C and its control at 43°C and 28 °C.

As mentioned above, controls were carried out at both 43°C and 28°C to compare the effect of heat alone on insects. In Table 1, the effect of 43°C alone in the control of both insects was apparent. Higher mortalities were observed in the controls at 43°C compared to the controls at 28°C. The adult stage of *O. surinamensis* was the most tolerant one with 99.5 % mortality on average. Larval and pupal stages could not survive the treatment and 100% mortality was recorded (Table 1).

Although some eggs of both species hatched, they could not survive more than 24 h after the treatment. Average egg mortalities were 97.5% for *T. castaneum* and 98% for *O. surinamensis*. Therefore, the mortalities for both species were considered as 100%.

### Discussion

The target temperature of 43°C, as is shown in Fig. 1, was reached in less than 30 min in the jars. In practice, when handling large commercial volumes, this process may take several hours or days to reach the target temperature at the core of the stack/bags, depending on the volume size (Donahaye et al., 1995).

According to Navarro (2012), to obtain rapid killing when using  $N_2$ , the  $O_2$  concentration must be lower than 1%. Timlick et al. (2002) reported that in a sealed commercial storage that maintained less than 1%  $O_2$ , insect mortality was completed after 14 d at 17°C. Therefore,  $N_2$  has been considered unsuitable for bulk commodity treatment at export locations due to the length of time required for 100% mortality.

However, insect mortality increases more rapidly as temperatures rise (Navarro, 2012). In heat treatments carried out in commercial facilities, it was found that the young larvae of *T. castaneum* were the most heat tolerant at 50 to  $60^{\circ}$ C within 24 to 36 h (Subramanyam et al., 2011). In our study, the effect of high temperature (43°C) in the control is more pronounced especially for the egg, larva and pupa of *T. castaneum* which were much more susceptible than the adult (Table 1). In the control of *O. surinamensis*, all development stages were affected by the high temperature. Although eggs hatched to larvae, they were dead one day later.

After exposure to 43°C in the control, adult *Tribolium castaneum* was the most tolerant stage to this high temperature (<2% mortality), followed by the pupal stage (42% mortality, Table 1). Navarro (1978) reported the significant differences of adult mortality of *T. castaneum* in N<sub>2</sub> which ranged between 2 d to 4.5 d at 0.1 to 1.0% O<sub>2</sub> at 26°C. However, in these trials, adults of *T. castaneum* at 1% O<sub>2</sub> were much more susceptible due to the high temperatures. This is consistent with Sakka et al. (2020) that a complete mortality was achieved in 2.5 d at 40°C and 1% O<sub>2</sub> and was emphasized at 28°C where they obtained mean mortality of 96.7% after 9 d of exposure time.

The pupae and larvae of both species were more susceptible to the treatment than the adults and eggs in both species. Although it is reported that some percentage of eggs did hatch (Fig. 3), the hatched larvae were found all dead immediately after the treatment. The reason for the tolerance of eggs to the treatment might be due to the low respiration rate of the eggs, but the complete control was achieved due to the high temperatures on the hatched eggs which was lethal due to their susceptibility. Fields (2002) reports that at a temperature range of 45-50°C, death is achieved in less than 24 h. However, at a range of 42-45°C there is no data.

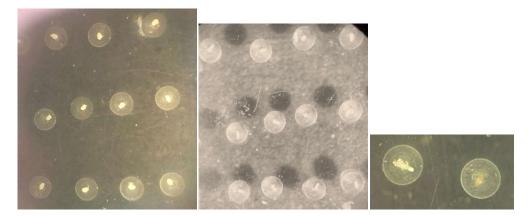


Fig. 3: *T. castaneum* eggs after treatment (left), *O. surinamensis* eggs after treatment (middle), and hatched eggs (right).

Protection of the beneficial qualities of cereals during storage depends on many factors. Among the detrimental factors that reduce the quality of cereals are insects and microflora (Navarro and Donahaye, 2005). Although the main objective of modified or controlled atmospheres is to control insect pests, the use of it enables quality preservation of the product as well, due to the sufficient sealed structure which maintains the vapor pressure (Navarro and Navarro, 2018). Even though in these trials, quality tests were not carried out, it is assumed that no quality deterioration occurred and the organoleptic characteristics were not affected.

#### Conclusions

Treating both stored product insects, *T. castaneum* and *O. surinamensis* at 43°C in 1% O<sub>2</sub> using N<sub>2</sub> for 48 h resulted in very effective treatment. The survival rate was less than 0.5 % of adult of *O. surinamensis*. Increased mortality was observed at 43°C compared to 28°C. The combination of 43°C with 1% O<sub>2</sub> in N<sub>2</sub> atmosphere was demonstrated as very effective in the control of all stages of both species. More data should be obtained to fill in the gaps of knowledge as for the tolerant stages of other insect species.

# References

- Andrews AS, Annis P, Newman CR (1994) Sealed storage technology for Australian farms. Pp. 27-36. In: Highley E, Wright EJ, Banks HJ, Champ BR (eds) Proc 6th Intern Working Conf Stored-Prod Prot, Canberra, Australia.
- Annis PC, van Graver SJ (1991) Suggested recommendations for the fumigation of grain in the ASEAN region. Part 2. Carbon dioxide fumigation of bag-stacks sealed in plastic enclosures: An operations manual. ASEAN Food Handling Bureau Kuala Lumpur, Malaysia and ACIAR, Canberra, Australia, 58 p.
- Athanassiou CG, Chiou A, Rumbos CI, Sotiroudas V, Sakka M, Nikolidaki EK, Panagopoulou EA, Kouvelas A, Katechaki E, Karathanos VT (2017) Effect of nitrogen in combination with elevated temperatures on insects, microbes and organoleptic characteristics of stored currants. J Pest Sci **90**: 557-567.

- Banks J, Fields P (1995) Physical methods for insect control in stored-grain ecosystems. Pp. 353-409. In: Jayas DS, White NDG, Muir WE (eds) Stored Grain Ecosystems. Marcel Dekker, New York.
- Donahaye EJ (2000) Current status of non-residual control methods against stored product pests. Crop Prot **19**: 571-576.
- Donahaye EJ, Navarro S, Rindner M (1995) The influence of low temperatures on three stored-product insects. J Stored Prod Res **31**: 63-70.
- Fields P, White N (2002) Alternatives to methyl bromide treatments for stored-product and quarantine insects. Annual Review of Entomol **47**: 331-359.
- Navarro S (1978) The effects of low oxygen tensions on three stored product insect pests. Phytoparasitica **6**: 51-58.
- Navarro S (2006) Modified atmospheres for the control of stored product insects and mites. Pp. 105–146. In: Heaps JW (ed) Insect Management for Food Storage and Processing, 2nd edn. AACC International, St. Paul.
- Navarro S (1999) Pressure tests for gaseous applications in sealed storages: Theory and practice. Pp. 385-390. In: Zuxun J, Quan L, Yongsheng L, Xianchang T, Lianghua G (eds) Proc 7<sup>th</sup> Intern Working Conf Stored-Prod Prot, Beijing, China.
- Navarro S (2012) The use of modified and controlled atmospheres for the disinfestation of stored products. J Pest Sci **85**(3): 301-322.
- Navarro S, Donahaye E (2005) Innovative environmentally friendly technologies to maintain quality of durable agricultural produce. Pp. 205-262. In: Ben-Yehoshua S (ed) Environmentally Friendly Technologies for Agricultural Produce Quality, CRC Press, Taylor & Francis Group, Boca Raton, FL.
- Navarro S, Navarro H (2018) The significance of vapor pressure in quality preservation of stored commodities under gastight conditions. Julius-Kühn-Archiv **463**: 549-556.
- Navarro S, Donahaye E (1990) Generation and application of modified atmospheres and fumigants for the control of storage insects. Pp. 152-165. In: Champ BR, Highley E, Banks HJ (eds) Proc Intern Conf Fumi and Contr Atm Storage of Grain. ACIAR Proc. 25. Brown Prior Anderson Pty. Ltd., Burwood, Victoria, Australia.
- Thai HN (2012) The risk of becoming the cashew processing factory of the world. Vietnam Cashew Association: Ho Chi Minh City.
- Sakka MK, Gatzali F, Karathanos VT and Athanassiou CG (2020) Effect of nitrogen on phosphinesusceptible and -resistant populations of stored product insects. Insects 11: 885.
- Subramanyam B, Mahroof R, Brijwani M (2011) Heat treatment of grain-processing facilities for insect management: a historical overview and recent advances. Stewart Postharvest Review **3**: 1.
- Timlick B, Dickie G, McKinnon D (2002) Nitrogen as a major component of a controlled atmosphere to manage stored product insect pests in large vertical storage. Integrated Protection of Stored Products, IOBC Bulletin 25; **3**: 193-197.